

S0040-4039(96)00142-6

A Novel Convenient Route to the Naturally Occurring 3-Oxoacyl-L-Homoserinelactones and Related Bacterial Autoinducers.

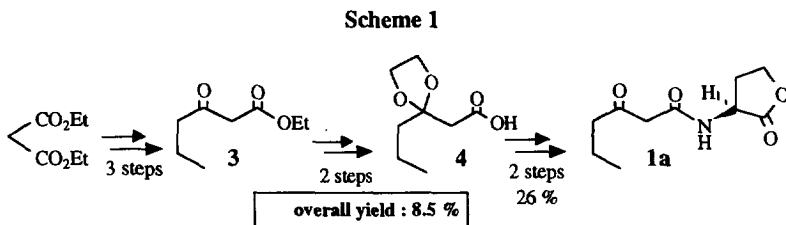
Mouloud Dekhane*, Kenneth T. Douglas and Peter Gilbert

Department of Pharmacy, University of Manchester, Manchester, M13 9PL, U.K.

Abstract : The naturally occurring 3-oxohexanoyl-L-homoserinelactone (**1a**), a bacterial autoinducer, has been prepared in 47 % overall yield by condensing stable 3-oxohexanoic acid (**2**), prepared by hydrolysis from the corresponding ester (**3**), with L-homoserinelactone using hydroxybenzotriazole (HOBT) and dicyclohexylcarbodiimide (DCC) in non-aqueous media.

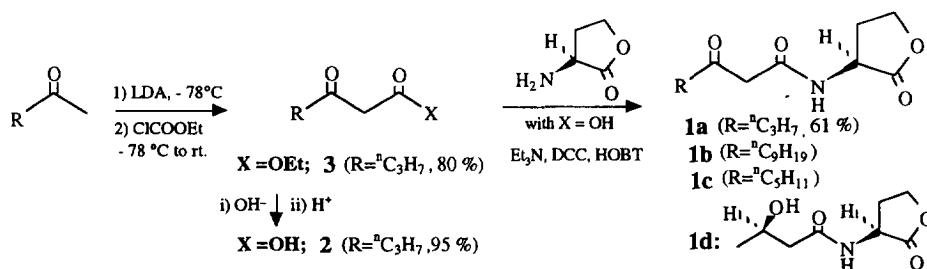
Analogues of homoserinelactone (HSL) have recently been shown to have a major role in cell-cell communication.¹ HSLs are produced during the normal growth of many Gram-negative bacteria, and at high cell densities, reach a threshold concentration for bacterial detection. This leads to global regulation of a variety of physiological processes.^{1,2} HSLs isolated to date are : (**1a**),^{3,4} (**1b**),⁵ (**1c**)⁶ and (**1d**)⁷ (Scheme 2). Most microbiological studies to date have isolated HSLs from batch fermentation but there is a need for a rapid, simple synthetic method, versatile to allow for ready synthesis of variants.

There are currently three reported syntheses of N-substituted-L-homoserinelactones of the autoinducer classes. The first reported, for N-(3-oxohexanoyl)-L-homoserinelactone (**1a**), gave the racemate and it was not possible to calculate the overall yield.³ A range of analogues has been reported using either carbodiimide or acyl chloride activation of the protected 3-ketoacid. The chirally pure product was synthesized (Scheme 1), in 26 % yield for the two steps from the ethylene glycol ketal of 3-oxohexanoic acid **4**, using a water-soluble carbodiimide.⁶ Recently, N-(3-oxododecanoyl)-L-homoserinelactone (**1b**) was synthesized similarly.⁵ In these routes β -ketoester **3** was prepared (three steps) from diethyl malonate ⁸ and the 3-oxo function ketal-protected (Scheme 1).



We now report a route to this family of homoserinelactone autoinducers (Scheme 2) which avoids the need for these protection and deprotection steps.

Scheme 2



Previous studies differ from the synthetic route in Scheme 2 mainly in starting with diethyl malonate, 3-oxoprotection and an aqueous environment for condensation of the 3-oxoacid with L-homoserine lactone. We have found that by using hydroxybenzotriazole and dicyclohexylcarbodiimide in nonaqueous media there is no need to protect the 3-oxo site, leading to an improved overall yield with fewer steps. Ethyl 3-oxohexanoate **3** was condensed with ethyl chloroformate by means of lithium diisopropylamide in 80 % yield and base hydrolysis of this to the free acid **2** was carried out in 95 % yield. Although the β -ketoacid **2** has been reported as unstable at room temperature,⁹ the spectral data for **2** were entirely consistent with the β -ketoacid structure. Finally, 3-oxo-N-(tetrahydro-2-oxo-3-furanyl)hexanamide (**1a**) was prepared in 61 % yield by reaction of **2** with L-homoserinelactone in dry dichloromethane with activation by DCC and HOBT.

In conclusion, a new, short, high-yielding route to the biologically interesting and useful synthetic N-(3-oxohexanoyl)-L-homoserinelactone (**1a**) has been developed. This synthetic methodology is clearly readily applicable to other analogues.

ACKNOWLEDGEMENT

We are grateful to the BBSRC for financial support under a ROPA award.

REFERENCES

- Williams, P. A.; Bainton, N. J.; Swift, S.; Chabra, S. R.; Winson, M. K.; Stewart, G. S. A. B.; Salmond, G. P. C.; Bycroft, B. W. *FEMS Microbiology Lett.* **1992**, *100*, 161-168
- Gambello, M. J.; Kaye, S.; Iglewski, B. H., *Infection and Immunity* **1993**, *61*, 1180-1184
- Eberhard, A.; Burlingame, A. L.; Eberhard, C.; Kenyon, G. L.; Nealson, K. H.; Oppenheimer, N. J. *Biochemistry* **1981**, *20*, 2444-2449.
- Bainton, N. J.; Stead P.; Chabra, S. R.; Bycroft, B. W.; Salmond, G. P. C.; Stewart, G. S. A. B.; Williams, P. *Biochem. J.* **1992**, *288*, 997-1004
- Pearson, J. M.; Gray, K. M.; Passador, L.; Tucker, K. D. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 197-201
- Zhang, L.; Murphy, P. J.; Kerr, A.; Tate, M. *Nature (London)* **1993**, *362*, 446-448
- Cao, J.-G.; Meighen, E. A. *J. Biol. Chem.* **1989**, *264*, 21670-76.; *J. Bacteriol.* **1993**, *175*, 3856-3862
- Wierenga, W.; Skulnick, I. *J. Org. Chem.* **1979**, *44*, 310-311
- Lewis, N.; Mander, L. N.; Sethi, P. *Tetrahedron Lett.* **1983**, *48*, 5425-5428. Koch, G. K.; Kop, J. M. M. *Tetrahedron Lett.* **1974**, *7*, 603-606

(Received in UK 11 December 1995; revised 18 January 1996; accepted 26 January 1996)